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1-Phenylcarbamoyl-2-pyrazolines: a New Class of Insecticides. 1. Synthesis and Insecticidal Properties of 3-Phenyl-1-phenylcarbamoyl-2-pyrazolines

Kobus Wellinga, Arnold C. Grosscurt, and Roelof van Hes*

Upon the discovery of the powerful insecticidal properties of 3-(4-chlorophenyl)-1-(4-chlorophenylcarbamoyl)-2-pyrazoline (PH 60-41), a large number of analogues was prepared. The structures of these compounds were confirmed by NMR. Their insecticidal properties were evaluated with larval stages of *Aëdes aegypti* L., *Pieris brassicae* L., and *Leptinotarsa decemlineata* Say. Many of these compounds proved to be excellent insecticides, bringing about the same symptoms as PH 60-41. From the point of view of biological activity and of economic aspects, PH 60-41 appeared to be the most promising compound for several fields of application.

In a recent report from our laboratories (Mulder et al., (1975), the new insecticidal compound 3-(4-chlorophenyl)-1-(4-chlorophenylcarbamoyl)-2-pyrazoline (PH 60-41) was introduced. This compound originated from a research program with derivatives of 3-aryl-2-pyrazolines. On larvae and adults of several insect species this compound proved to be a powerful stomach toxicant with some contact activity. Within 24 h after application, susceptible insects showed convulsions. They lost their grip on the leaves, fell off the plants, and finally died.

In the present paper we report the synthesis of a series of 3-phenyl-1-phenylcarbamoyl-2-pyrazolines and their evaluation as potential insecticides. Parts 2 and 3 of this series of papers (van Hes et al., 1977; Grosscurt et al., 1978) will show that substitution in the 2-pyrazoline nucleus of two phenyl rings at the 3,5 and 3,4 positions, respectively, instead of one at the 3 position gives rise to products with insecticidal properties of the same order or even better. CHEMICAL METHODS

Microanalyses were carried out in the Analytical Department of the Institute for Organic Chemistry TNO, Utrecht, Netherlands, under the supervision of W. J. Buis. The carbon-13 spectra were measured with a Bruker WH 270 spectrometer, operating at 67.89 MHz. Nuclear magnetic resonance spectra were recorded on a Varian HA 100 spectrometer. Chemical shifts were measured with tetramethylsilane as the internal reference and with deuteriodimethyl sulfoxide as a solvent. The melting points are uncorrected.

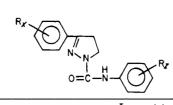
One general method was used for the preparation of the various compounds mentioned in Tables I and II. They were prepared according to Scheme I.

4'-Chloro-3-dimethylaminopropiophenone Hydrochloride. A suspension of 30.9 g of 4-chloroacetophenone (0.2 mol), 21.2 g of dimethylamine hydrochloride (0.26 mol), and 7.8 g of paraformaldehyde (0.26 mol) in a mixture of 32 mL of ethanol and 0.4 mL of concentrated hydrochloric acid was refluxed for 2 h. After cooling, 160 mL of acetone was added. The crystals formed were collected, washed with acetone, and dried. Yield 35.8 g (72%), mp 173-175 °C [literature (Nobles, 1958), 176 °C].

3-(4-Chlorophenyl)-2-pyrazoline. A warm solution of 24.8 g of 4'-chloro-3-dimethylaminopropiophenone hydrochloride (0.1 mol) in 70 mL of methanol was added in 10 min to a mixture of 14 mL of hydrazine hydrate, 7.2 mL of 50% sodium hydroxide, and 18 mL of methanol.

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Table I.^a Insecticidal Activities of



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			Lowest tes		n with $\geq 90\%$ m	ortality
Compd no.	R _x	Rz	Leptinotarsa decemlineata Say	Pieris y brassicae L.	Aëdes aegypti L.	Mp, °C
		a. R_x	= H, alkyl			
1	Н	Н	100	>300	>1	160
2	Н	$4 \cdot n \cdot C_{3}H$	10	300	1	120
3	Н	$4 - t - C_4 H_9$	10	300	>1	145
4	Н	4-F	30	>300	>1	137
5	Н	4-Cl	3	>300	1	148
6	H	3,4-Cl ₂	10	>300	>1	168
7	4-CH,	4-CH,	30	>300	1	159
8	4-CH,	4-Cl	30	>300	0.3	139
9	4-C₂H ₅	4-Cl	30	100	>1	116
10	$4-C_2H_5$	$4-SO_2C_2H_5$	30	300	>1	172
11	$4 \cdot n \cdot C_3 H_3$	4-Cl	100	>300	>1	153
12	$4 - i - C_3 H$	4-Cl	10	100	>1	131
13	$4 - i - C_3 H_3$	$4-SO_2C_2H_5$	300	>300	>1	195
14	$4 \cdot n \cdot C_4 H_{2}$	4-C₂H₅ 4-Cl	300 30	100	1	147
15	$4 - t - C_4 H_9$			300	>1	150
	a a		gen; $R_z = H$, alkyl			. .
16	2-Cl	$4 \cdot n \cdot C_3 H_7$	100	>300	>1	87
17	4-Cl	H	30	>300	>1	160
18^b	4-Cl	2-CH,	>300	> 300	>1	162
19	4-Cl	4-CH,	30	>300	>1	182
20	4-Cl	$4 - C_2 H_5$	10	>300	>1	140
21	4-C1	$4 - n - C_3 H_7$	3	10	0.3	151
22	4-Cl	$4 - i - C, H_7$	1	10	>1	143
23	4-Cl	$4 \cdot n \cdot C_4 H_9$	30	30	>1	134
24	4-Cl	$4 - i - C_4 H_9$	30	10	>1	153
25	4-Cl	$4 - t - C_4 H_9$	3	30	>1	162
26 27 ⁶	4-Cl	$4 \cdot n \cdot C_8 H_{17}$	100	>300	>1	107
27 ⁶ 28 ^b	4-Cl	$4 - n - C_{12} H_{25}$	>300	>300	>1	87
	$2,4-Cl_{2}$	4-CH,	> 300	>300	>1	123
29 30	4-Br 4-Br	H 4- <i>n</i> - C_3H_7	100 10	>300 100	>1 0.3	$\begin{array}{c} 155 \\ 167 \end{array}$
30	4-DI			100	0.0	107
0.1	4 17	c. $R_x =$	$R_z = halogen$	> 200	1	150
31	4-F	4-Cl	3	>300	1	150
32	4-F	3,4-Cl	30	> 300	>1	162
33	2-Cl	4-Cl	100	> 300	>1	123
34	2-Cl	3,4-Cl	100	>300	>1	144
35	3-Cl	4-Cl	30	>300	1	161
36	3-Cl	3,4-Cl ₂	100	>300	>1	$180 \\ 178$
37 38 ⁶	4-Cl 4-Cl	4-F 2-Cl	10 >300	>300 >300	>1 >1	$\begin{array}{c} 173 \\ 175 \end{array}$
39		2-Cl 3-Cl				147
40	4-Cl	4-Cl	100 3	>300 100	>1 0.3	178
41	4-Cl 4-Cl	2,4-Cl ₂	100	>300	>1	168
41 42 ^b	4-Cl	2,4-Cl ₂ 2,5-Cl ₂	>300	>300	>1	240
43	4-Cl	3,4-Cl ₂	10	300	>1	187
44 ^b	4-Cl	3,5-Cl,	> 300	>300	>1	202
45	4-Cl	2,4,5-Cl ₃	>300	>300	>1	227
46	4-Cl	2,4,5-Cl ₃	> 300	> 300	>1	200
47	4-Cl	4-Br	10	100	0.3	181
48	4-Cl	4-I	10	30	0.3	162
40	3,4-Cl ₂	4-Cl	10	>300	1	180
45 50	3,5-Cl ₂	4-Cl	300	>300	>1	200
51	4-Br	4-Cl	3	100	0.3	144
52	4-Br	3,4-Cl ₂	10	300	>1	192
53	4-Br	4-Br	3	30	>1	147
54	4-I	4-Cl	10	300	0.3	165
		/				
55	2-C1	d. $R_x = halogen$; $R_z = 4 \cdot O \cdot i \cdot C_3 H_7$	= electron-donatin 100	sig groups >300	>1	93
56	4-Cl	4-CH ₃ , 3-Cl	30	>300	>1	189
	4-Cl	$4-O(1_3, 3-O(1_3)_2)$	100	>300	>1	163
57		4 0011	3	>300	>1	135
57 58	4-C1					
58	4-Cl 4-Cl	4-OCH, 4-OC-H.		300		
	4-Cl 4-Cl 4-Cl	4-OC ₂ H ₅ 4-OC ₂ H ₅ 4-O- <i>i</i> -C ₃ H ₇	10 3	300 10	1 0.3	139 130

			Lowest test concentration with $\ge 90\%$ mortality				
Compd no.	R_x	R _z	Leptinotarsa decemlineata Say	Pieris brassicae L.	Aëdes aegypti L.	Mp, °C	
62	4-Cl	4-OCH,, 3-Cl	300	>300	>1	208	
		e. $R_x = halogen; R_z$	= electron-attractin	g groups			
63	4-Cl	3-CF,	10	>300	>1	138	
64	4-Cl	4-CF ₃	1	10	0.1	120	
65	4-Cl	3-NO,	100	>300	>1	210	
66	4-Cl	4-NO ₂	10	>300	>1	225	
67	4-Cl	4-COĆH,	300	>300	>1	174	
68	4-Cl	4-CN	3	>300	>1	210	
6 9	4-Cl	4-SO ₂ CH,	30	>300	>1	225	
70	4-Cl	4-SO ₂ C ₂ H ₅	10	100	² >1	187	
71	4-Cl	$4 \cdot SO_2 \cdot n \cdot C_4 H_9$	30	100	>1	146	
72	4-B r	4-CN ²	10	>300	>1	227	
		f. $R_r = elect$	ron donating group	5			
73	$4 - N(CH_{3})_{2}$	4-Cl	3 1	>300	1	189	
74	$4 - N(CH_{3})_{2}$	$4 - N(CH_3)_2$	300	>300	>1	209	
75	3-OCH,	4-Cl	30	>300	>1	129	
76	3-OCH,	3,4-Cl,	100	>300	>1	136	
77	4-OCH	4-Cl	10	>300	1	132	
78	4-OCH	$3, 4 - Cl_2$	30	>300	>1	170	
79	4-0- <i>i</i> -C ₃ H ₇	4-Cl	10	30	0.3	141	
80	4-SCH,	4-Cl	30	>300	1	154	
		g. R_x = electr	on attracting group	s			
81	4-C ₆ H ₅	4-Čl	30 1	>300	>1	290	
82	3-NO ₂	4-Cl	30	>300	>1	214	
83	3-NO ₂	3,4-Cl,	300	>300	>1	210	
84	$4-SO_2CH_3$	4-Cl	300	>300	>1	246	

^a Applied concentrations, in parts per million: *Aëdes aegypti* L., 1, 0.3, 0.1, 0.03, etc.; *Pieris brassicae* L., 300, 100, 30, 10, etc.; *Leptinotarsa decemlineata* Say, 300, 100, 30, 10, etc. ^b These compounds are slightly active on *Leptinotarsa decemlineata* Say and/or on *Pieris brassicae* L. at a concentration higher than 300 ppm.

Table II.^a Insecticidal Activities of



Compd no. R ₁			R,	Lowest test concentration with $\geq 90\%$ mort.				
	X R ₂	R_2		Leptinotarsa decem- lineata Say	Pieris brassicae L.	Aëdes aegypti L.	Mp, °C	
1	4-ClC ₆ H ₄	0	CH ₃	4-ClC ₆ H ₄	10	300	>1	146
2	$4 - ClC_6H_4$	0	C ₂ H ₅	4-ClC ₆ H₄	10	300	>1	133
3	$4-ClC_6H_4$	0	$n - C_3 H_7$	$4-ClC_{6}H_{4}$	100	300	>1	132
4	CH,	0	Н	$4 - ClC_6 H_4$	>300	>300	>1	138
5	n-C₄H,	0	Н	4-ClC ₆ H₄	>300	>300	>1	123
6	$c - C_{6} H_{11}$	0	Н	4-CIC°H₄	100	>300	>1	156
7	$n - C_{11} H_{23}$	0	Н	4-ClC ₆ H ₄	>300	>300	>1	71
8	4-ClC ₆ H ₄	0	Н	$n-C_3H_7$	>300	>300	>1	90
9	4-ClC ₆ H₄	\mathbf{s}	Н	4-CIC, H	30	>300	>1	198

^a Applied concentrations, in parts per million: *Aëdes aegypti* L., 1, 0.3, 0.1, 0.03, etc.; *Pieris brassicae* L., 300, 100, 30, 10, etc.; *Leptinotarsa decemlineata* Say, 300, 100, 30, 10, etc.

After refluxing for 45 min, the methanol was distilled off at reduced pressure. The residue was dissolved in dichloromethane and washed with water. Evaporation of the solvent gave the crude 2-pyrazoline (16.0 g), which could be used without further purification. The 2-pyrazolines of this type were not very stable and had to be stored under nitrogen in the refrigerator.

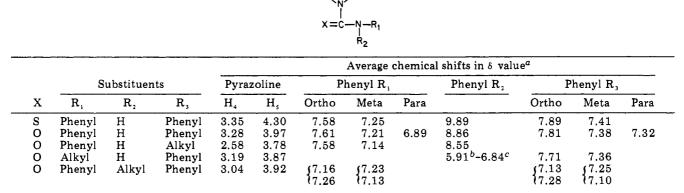
3-(4-Chlorophenyl)-1-(4-chlorophenylcarbamoyl)-2-pyrazoline [Table I, Compound 40 (I,40); PH 60-41]. To a suspension of 5.4 g of 3-(4-chlorophenyl)-2-pyrazoline (0.03 mol) in 100 mL of dry ether, three drops of triethylamine and 4.6 g of 4-chlorophenylisocyanate (0.03 mol) were added. The solution became clear and a precipitate of the reaction product appeared. After stirring for 1.5 h, the precipitate was collected and dried, yielding 7.5 g (75%) of compound 40, mp 176–178 °C.

Anal. Calcd for $C_{16}H_{13}Cl_2N_3O$ (mol wt, 334.20): C, 57.50; H, 3.92; Cl, 21.22; N, 12.57. Found: C, 57.2; H, 3.9; Cl, 21.4; N, 12.6.

3-(4-Chlorophenyl)-1-[N-(4-chlorophenyl)-Nmethyl]carbamoyl-2-pyrazoline (II,1).

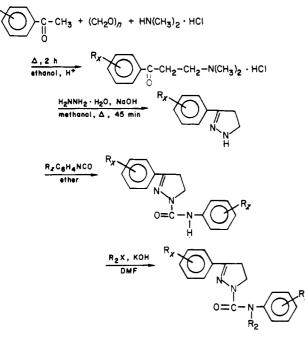
To a stirred suspension of 3.3 g of 3-(4-chlorophenyl)-1-(4-chlorophenylcarbamoyl)-2-pyrazoline (= I,40; 0.01 mol) and 1 g of potassium carbonate in 20 mL of DMF,

Table III. Proton NMR Data of



^a The pyrazoline protons appear as broad triplets with J = 11 Hz. Actually it is an AA'XX' spin system with $J_{AX} + J_{AX'} = 22$ Hz and $J_{AX} - J_{AX'} < 7$ Hz. The standard deviation from the average chemical shifts of the pyrazoline protons was 0.03. The chemical shifts of the phenyl and NH protons are calculated for the unsubstituted derivatives, with the help of the aromatic substituent shifts of Table IV. The standard deviation for the aromatic protons was 0.05 and for the NH protons 0.07. ^b Cyclohexyl. ^cn-alkyl.

Scheme I. Preparation of Substituted 3-Phenyl-1phenylcarbamoyl-2-pyrazolines



0.7 g of powdered potassium hydroxide (0.0125 mol) was added at 0 °C. After 15 min 0.7 mL of methyl iodide was added. The mixture was stirred for 0.5 h at 0 °C, then for 1.5 h at room temperature, and finally poured into ice water. The precipitate was collected, dried, and washed with petroleum ether. Yield 2.8 g (80%), mp 144-146 °C.

Anal. Calcd for $C_{17}H_{15}Cl_2N_3O$ (mol wt, 348.25): C, 58.63; H, 4.34; Cl, 20.37; N, 12.07. Found: C, 58.4; H, 4.4; Cl, 20.4; N, 12.0.

The compounds 4-7 and 9 of Table II were prepared according to the procedure described for compound I,40.

1-(4-Chlorophenylcarbamoyl)-3-n-propyl-2pyrazoline (II,8). 3-n-Propyl-2-pyrazoline was prepared according to procedures described in the literature (Kossanyi, 1965; Smith, 1952). The compound II,8 was prepared from 3-n-propyl-2-pyrazoline according to the procedure described for compound I,40; mp 88-90 °C.

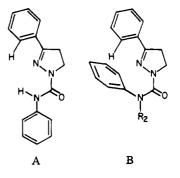
Anal. Calcd for $C_{13}H_{16}ClN_3O$ (mol wt, 265.75): Cl, 13.34; N, 15.82. Found: Cl, 13.4; N, 15.7. The following compound has been prepared for comparison with compound I,40 (see Results and Discussion section).

3-(4-Chlorophenyl)-1-(4-chlorophenylcarbamoyl)pyrazole. The synthesis of 3-(4-chlorophenyl)pyrazole is described in the literature (Kochetkov, 1958). To a solution of 1.96 g of 3-(4-chlorophenyl)pyrazole (0.011 mol) in 25 mL of acetone, 1.7 g of 4-chlorophenylisocyanate (0.011 mol) and three drops of triethylamine were added. After refluxing for 1 h, the reaction mixture was allowed to stand at room temperature for 24 h. The crystals were collected, washed with ether, and dried, yielding 2.3 g (63%): mp 149–150 °C; NMR (Me₂SO) δ 7.13 (1, d, J = 2.6 Hz, N-CH=CH), 8.49 (1, J = 2.6 Hz, NCH=CH), 8.11 (2, d, 1-phenyl), 7.56 (2, d, 1-phenyl), 7.85 (2, d, 3-phenyl), 7.45 (2, d, 3-phenyl), 10.43 (1, S, NH).

Anal. Calcd for $C_{16}H_{11}Cl_2N_3O$ (mol wt, 332.20): C, 57.85; H, 3.34; Cl, 21.35; N, 12.65. Found: C, 57.9; H, 3.3; Cl, 21.5; N, 12.6.

NMR SPECTRA

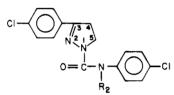
The proton NMR data of the 3-phenyl-1-phenylcarbamoyl-2-pyrazolines are presented in Tables III and IV. All the chemical shifts accord with expectations, except those of the aromatic protons of the N-alkyl (R_2 = alkyl) derivatives. Particularly the shifts of the ortho protons of the 3-phenyl ring are too low. However, the ¹³C chemical shifts of these N-alkyl derivatives are normal, compared with those of the corresponding NH derivatives (Table V). The anomalous chemical shifts must therefore be due to steric effects rather than to effects through the bonds. It may be assumed that the main conformation of the NH derivatives is that of structure A. A great con-



			Prot	on		
		Aromatic proton	S		φ -NH proton ^a	
	·····		Position of s	ubstituent	· · · · · · · · · · · · · · · · · · ·	
Substituent	Ortho	Meta	Para	Ortho	Meta	Para
-NO,	$+0.94^{a}$	+0.27	$+0.34^{a}$		+0.59	+0.61
-COĆH,	$+0.67^{a}$	$+0.26^{a}$				+0.34
-SO,R	$+ 0.59^{a}$	$+0.29^{a}$				+0.46
-CN ²	+0.49	+0.24				+0.53
-F	-0.13^{a}	$+0.07^{a}$		-0.34		+ 0.11
-Cl	+0.10	+0.07	+0.03	-0.20	+0.25	+0.15
-Br	+0.24	+0.02				+0.17
-I	+0.38	-0.07^{a}				+0.16
-CF ₃	+ 0.39	+ 0.29	$+0.36^{a}$		+0.42	+0.42
-C (+ 0.03	-0.05				-0.08
-ĆH	-0.06	-0.02				-0.08
-CH,-	-0.13	-0.07				-0.08
-CH,	-0.17	-0.07		- 0.48		-0.08
-OR	-0.41	-0.04				-0.10
-SCH,	-0.05^{a}	0^a				+0.06
-NR ₂	-0.67	-0.17				-0.25

^a Data are calculated from 3-phenyl-1-phenylcarbamoyl-2-pyrazolines, 3,4-diphenyl-1-phenylcarbamoyl-2-pyrazolines (Grosscurt et al., 1978), 3,5-diphenyl-1-phenylcarbamoyl-2-pyrazolines (van Hes et al., 1977); other data were obtained from the literature (Gove, 1973).

Table V. ¹³C NMR Chemical Shifts of



			D –
Carbon atom	s	$R_2 = H$	$\begin{array}{c} \mathbf{R}_{2} = \\ n \cdot \mathbf{C}_{3} \mathbf{H}_{7} \end{array}$
Pyrazoline ring		151.7 [°] 31.6	$\begin{array}{r}151.4\\30.2\end{array}$
1-Phenylcarbamoyl	C _s C=O	$\begin{array}{r} 44.8\\152.7\\\end{array}$	46.6 154.7
	$ \begin{array}{c} \mathbf{C}_{1}\\ \mathbf{C}_{2}\\ \mathbf{C}_{3}\\ \mathbf{C}_{4}\\ \mathbf{C}_{1} \end{array} $	$138.4 \\ 120.8 \\ 128.2$	$143.9 \\ 127.5 \\ 127.9$
3-Phenvl	C₄ C₄	126.0 130.3	129.2 130.6
		$128.6 \\ 128.3$	$128.53 \\ 128.45$
	C_4° α -CH ₂	134.6	$\begin{array}{r}134.1\\52.4\end{array}$
$\mathbf{R}_2 = n \cdot \mathbf{C}_3 \mathbf{H}_7$	β -CH ₂ γ -CH ₃		$\begin{array}{c} 21.0\\ 11.0 \end{array}$

tribution of the conformation of structure B can for the N-alkyl derivatives bring about the strong deshielding of the ortho protons of the 3-phenyl ring. The stronger deshielding of the C_4 protons compared with the C_5 protons of the pyrazoline ring may also be due to this conformation.

BIOLOGICAL METHODS

The insecticidal evaluations were performed with suspensions made by pouring out, with stirring, quantities of a 1% stock solution in acetone into water. Compounds insufficiently soluble in acetone were thoroughly milled with water in a laboratory glass mill. Various species were tested as follows.

a. Aëdes aegypti L. (yellow fever mosquito). Aliquots of 100 mL of tap water containing 1, 0.3, 0.1, 0.03, etc. ppm, respectively, of the substituted 3-phenyl-1-phenyl-carbamoyl-2-pyrazolines were supplied with ten 1-day old

larvae and kept at 25 °C. The larvae were fed with a mixture of brewer's yeast and wheaten bread in a 1:1 ratio. After 14 days, when the pupae of the untreated insects had hatched, the mortality percentages were assessed with a correction for the natural mortality according to Abbott (1925). There were three replicates per treatment.

b. Pieris brassicae L. (large white butterfly). Potted cabbage seedlings were sprayed with a suspension of the toxicant until runoff. The suspensions contained 300, 100, 30, 10, 3, etc. ppm of the compound, respectively. When dry, the plants were placed in plastic cylinders, provided with five third-instar caterpillars and kept under a day/ night cycle of 18/6 h at a temperature and relative humidity of 24 °C, 60-70% and 19 °C, 80-90%, respectively.

There were three replicates per treatment. After 5 days the mortality percentages were calculated according to Abbott's formulae.

c. Leptinotarsa decemlineata Say (Colorado potato beetle). Potato shoots, placed in flasks containing tap water, were sprayed with a suspension of the compound until runoff. The concentrations of the suspensions were 300, 100, 30, 10, 3, etc. ppm, respectively. When dry, the shoots were placed in plastic cylinders and provided with ten third-instar larvae. After this, the procedure and the environmental circumstances were identical with those described for the large white butterfly.

RESULTS AND DISCUSSION

The overwhelming number of substitution possibilities in these compounds induced us to a drastic limitation in the synthetic work. Nevertheless, for reasons of efficiency, we shall in this article report merely on some of the compounds prepared thus far.

In Table I, subdivided in sections a, b, c, etc., the insecticidal properties are presented of the different types of substituted 3-phenyl-1-phenylcarbamoyl-2-pyrazolines. Table II includes the compounds with certain modifications in the molecule with regard to the basic structures.

Table Ia shows the results of the compounds with an unsubstituted 3-phenyl ring, "ring 3", and with those substituted with an alkyl substituent at the para position. The phenyl ring in the carbamoyl group, "ring 1", carries a somewhat miscellaneous substitution pattern. These compounds display a high level of activity on *Leptinotarsa*, particularly in the case of halogen and of (higher) alkyl substitution in the last mentioned ring (I: 5, 6, 12, 2). The activity of *Pieris* and *Aëdes* varies from moderate to negligible. Compounds 2 and 14 were active on all three test objects.

From Tables Ib to Ie, recording the test results of all compounds with a halogen substituent in ring 3 and with various substituents in ring 1, the following may be concluded. Para substitution in ring 1 appears to be favorable, compared with ortho and meta substitution (I: 19 vs. 18; 40 vs. 38 and 39; 64 vs. 63; 66 vs. 65), while with an increasing number of halogen atoms the activity vanishes entirely (I: 42, 44, 45, 46), the 3,4-dichloro compound 43, however, being an exception. Compound 41 is but slightly active. The same holds true for ring 3 (I: 40 vs. 33 and 35; 43 vs. 34 and 36, 21 vs. 16; 60 vs. 55) and again introduction of additional halogen atoms brings about a decrease in activity (I: 28, 50). Compound 49, the other 3,4-dichloro derivative, remains about as active as compound 40 = PH 60-41, the first representative of this class of insecticides, which has been described elsewhere by Mulder et al. (1975).

For optimal activity on Pieris, parahalogen in ring 3 combined with a paraalkyl substituent $>C_2H_5$ in ring 1 turns out to be essential (I: 21, 22, 23, 24, 25). A high activity level in this series is found with compounds 21, 22, and 25 on Leptinotarsa. Apart from the paraalkyl compounds, it appears from Table Id that derivatives with other electron-donating groups in ring 1 also yield good results with Leptinotarsa (I: 57, 58, 59, 60, 61). Even more pronounced are these results in case of electron-attracting groups, including halogen, in ring 1 (I: 31, 40, 48, 51, 53, 64, 68). The more outstanding compounds in both series are 21, 60 and 40, 48, 51, 64, respectively, which are active on the three test organisms involved. Strong electrondonating groups, when present in ring 3, give rise to derivatives with fair results in the case of Leptinotarsa (I: 73, 77, 79), while electron-attracting groups in this ring apart from halogen atoms, yield derivatives with rather moderate results, presented in Table Ig. Activity with Pieris and Aëdes in these series appears to be negligible, compound 79 being an exception.

With regard to Table Ic comprising compounds with halogen substituents in both rings, the following may be noted. Leptinotarsa responds rather similarly to the four possible 2-pyrazolines with a parachloro atom in ring 1 and with different parahalogens in ring 3 (I: 31, 40, 51, 54). This is the case also if the rings 1 and 3 change places in the 2-pyrazoline nucleus (I: 37, 40, 47, 48). In both cases the activity of the parafluoro derivatives is negligible on *Pieris* and *Aëdes* (I: 31, 37).

Several alterations in the basic molecule are detrimental

to insecticidal activity, as is exemplified in Table II. Replacement of ring 1 or of ring 3 by an alkyl group (II: 4, 5, 6, 7, and 8, respectively) or conversion of one of the outstanding compounds, PH 60-41 (I: 40), to its sulfur analogue (II: 9) reduces the activity considerably. PH 60-41 when converted to its 4,5-dehydro derivative, consequently the pyrazole analogue, appears to be practically inactive (not presented in the tables).

N-Alkylation in the carbamoyl group results also in a decrease in activity (II: 1, 2, 3).

At present the toxicological data, with respect to vertebrates, of only five substituted 3-phenyl-1-phenylcarbamoyl-2-pyrazolines have been obtained. When administered orally the LD_{50} values of these compounds toward mice vary from 1780 mg/kg (I: 60) to higher than 3160 mg/kg (I: 6, 21, 40, 48), while intraperitoneally these values vary from 422 mg/kg (I: 60), via 750 mg/kg (I: 21) and >1000 mg/kg (I: 48) to >3160 mg/kg (I: 6, 40). CONCLUSION

From the economic and practical aspects, 3-(4-chlorophenyl)-1-(4-chlorophenylcarbamoyl)-2-pyrazoline (I: 40; PH 60-41), which is active not only against the test objects involved but also toward a great number of other arthropods (Mulder et al., 1975), appears to be one of the most promising compounds in this series.

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Supplementary Material Available: A listing of microanalyses (1 page). Ordering information is given on any current masthead page.

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